



INTERNATIONAL APPLICATION PUBLISHED UNDER THE PATENT COOPERATION TREATY (PCT)

(51) International Patent Classification ⁵ : A61K 39/39, 30/00, 39/02	A1	(11) International Publication Number: WO 93/02706 (43) International Publication Date: 18 February 1993 (18.02.93)
(21) International Application Number: PCT/AU92/00377 (22) International Filing Date: 24 July 1992 (24.07.92) (30) Priority data: PK7457 26 July 1991 (26.07.91) AU (71) Applicant (for all designated States except US): COMMON-WEALTH SCIENTIFIC AND INDUSTRIAL RESEARCH ORGANISATION [AU/AU]; 14 Limestone Avenue, Campbell, ACT 2601 (AU). (72) Inventors; and (75) Inventors/Applicants (for US only) : REILLY, Wayne, Gerard [AU/AU]; 72 Briens Road, Northmead, NSW 2152 (AU). WHITTAKER, Robert, George [AU/AU]; 23 Ramsay Avenue, West Pymble, NSW 2073 (AU). JENNINGS, Philip, Anthony [AU/AU]; Commonwealth Scientific and Industrial Research Organisation, Division of Biomolecular Engineering, 103 Delhi Road, North Ryde, NSW 2113 (GB). FINNEY, Kenneth, Geoffrey [AU/AU]; 32 Willow Crescent, Ryde, NSW 2112 (AU).		(74) Agent: F.B. RICE & CO.; 28A Montague Street, Balmain, NSW 2041 (AU). (81) Designated States: AU, CA, JP, US, European patent (AT, BE, CH, DE, DK, ES, FR, GB, GR, IT, LU, MC, NL, SE). Published <i>With international search report.</i>
(54) Title: SELF-ADJUVANTING PEPTIDE VACCINE DELIVERY SYSTEM AND PRODUCTION THEREOF (57) Abstract <p>The present invention provides self-adjuvanting vaccines for use in raising antibodies to peptides without the use of oil or alum adjuvants. Further, the present invention provides methods of therapy using these vaccines and has particular application in chemical castration. In one aspect the vaccine comprises in admixture a peptide conjugated to 1-3 fatty acids and a peptide conjugated to a carrier protein. In preferred forms the peptide is conjugated to the fatty acids via a tromethamine or ethanolamine derivative. The preferred protein carrier is Type 4 fimbriae.</p>		

FOR THE PURPOSES OF INFORMATION ONLY

Codes used to identify States party to the PCT on the front pages of pamphlets publishing international applications under the PCT.

AT	Austria	FI	Finland	MN	Mongolia
AU	Australia	FR	France	MR	Mauritania
BB	Barbados	GA	Gabon	MW	Malawi
BE	Belgium	GB	United Kingdom	NL	Netherlands
BF	Burkina Faso	GN	Guinea	NO	Norway
BG	Bulgaria	GR	Greece	NZ	New Zealand
BJ	Benin	HU	Hungary	PL	Poland
BR	Brazil	IE	Ireland	PT	Portugal
CA	Canada	IT	Italy	RO	Romania
CF	Central African Republic	JP	Japan	RU	Russian Federation
CG	Congo	KP	Democratic People's Republic of Korea	SD	Sudan
CH	Switzerland	KR	Republic of Korea	SE	Sweden
CI	Côte d'Ivoire	LI	Liechtenstein	SK	Slovak Republic
CM	Cameroon	LK	Sri Lanka	SN	Senegal
CS	Czechoslovakia	LU	Luxembourg	SU	Soviet Union
CZ	Czech Republic	MC	Monaco	TD	Chad
DE	Germany	MG	Madagascar	TG	Togo
DK	Denmark	ML	Mali	UA	Ukraine
ES	Spain			US	United States of America

SELF-ADJUVANTING PEPTIDE VACCINE DELIVERY
SYSTEM AND PRODUCTION THEREOF

Background of the Invention

The present invention relates generally to
5 self-adjuvanting vaccines for use in raising antibodies to
peptides without the use of oil or alum adjuvants. The
present invention further relates to the use of this
vaccine and to methods of therapy. The present invention
has particular application in chemical castration.

10 The use of leutinising hormone releasing hormone
(LHRH) in chemical sterilisation of both male and female
animals is known. In prior art techniques animals are
immunised with preparations including LHRH and this
immunisation leads to dysfunction of the gonads and
15 consequent induced sterility. In male animals reduction
in testicular size relative to untreated controls may be
clearly seen. Unfortunately, the mere immunisation with
LHRH is not a totally effective method of chemical
castration. Typically, whilst in the majority of one
20 group of animals the immunisation with LHRH will have the
desired effect, the immunisation will have little effect
on a number of animals in that sample. As will be readily
appreciated this is a significant drawback because, for
example, if one unaffected male remains in the immunised
25 herd this male may fertilise a number of females within
the herd. For this reason chemical sterilisation
utilising LHRH has to date achieved only limited
application, with physical and other methods of
sterilisation still predominating.

30 In the present applicant's copending Australian
patent application No 17049/88 there is disclosed an
effective method of peptide production by protein
engineering. This method involves the culturing of
genetically engineered bacteria which produce, as
35 extracellular structures, Type 4 fimbriae, the peptide

being produced in association with these fimbriae. This method comprises culturing bacteria containing the gene encoding the fimbrial subunit of Type 4 fimbriae to which has been added at the C-terminal end the nucleic acid sequence encoding the desired peptide. In further work on this system the present inventors have found that not only does this method provide a simple and efficient means of producing a peptide but that the fimbrial protein acts as a powerful immunoadjuvant.

10 The present inventors have also produced a peptide having a sequence derived from LHRH in association with Type 4 fimbriae. It has been found that this peptide conjugated to bacterial Type 4 fimbriae provides a very effective chemical sterilisation agent.

15 In the present applicant's copending International Patent Application No PCT/AU90/00599 (the disclosure of which is incorporated herein by reference) there is disclosure of means of linking fatty acids and the like to peptides.

20 The present inventors have also found that co-administration of a peptide/fatty acid conjugate with a peptide/carrier protein (such as fimbrial protein) conjugate yields a heightened antibody response to the peptide.

25 In addition the present inventors have found that the antibody response in an animal can be enhanced by the use of the fimbrial protein of a Type 4 fimbriate bacteria.

Summary of the Invention

Accordingly, in a first aspect the present invention consists in a vaccine for use in raising an immune response to a peptide, the vaccine comprising in admixture the peptide conjugated to 1 to 3 fatty acids and the peptide conjugated to a carrier protein.

In a preferred embodiment of the first aspect of the present invention the peptide has or includes one of the

following amino acid sequences:-

SGGWSYGLRPGG;

WSYGLRP;

WSYGWLP; or

5 WSYGLQP.

In a second aspect the present invention consists in a vaccine for use in chemical sterilisation the vaccine comprising a peptide which has or includes an amino acid sequence selected from the group consisting of:-

10 SGGWSYGLRPGG, WSYGLRP, WSYGWLP and WSYLLQP,
the peptide being conjugated either to 1 to 3 fatty acids or to a fimbrial subunit.

In a preferred embodiment of the invention the carrier protein is a fimbrial sub unit protein, ovalbumin,
15 bovine serum albumin, tetanus toxin, or keyhole limpet haemocyanin. It is presently preferred, however, that the carrier protein is a fimbrial protein subunit and that the subunit proteins are assembled into fimbriae. It is most preferred that the fimbriae are Type 4 fimbriae.

20 The peptide is preferably conjugated to 1 to 3 fatty acids via a tromethamine or an ethanolamine derivative.

It is presently preferred that the peptide is linked to three fatty acids and more preferably that each are the same fatty acid. It is also preferred that the fatty acid
25 has a carbon chain of 3 to 18 carbon atoms and most preferably 16 carbon atoms.

The present inventors have also found that partial denaturation of the peptide/fimbrial protein carrier results in a higher antigenic response. This partial
30 denaturation is preferably obtained by treatment of the peptide/carrier protein at a pH of less than or equal to 4 and preferably at a pH of about 1. Given the enhanced response that this partial denaturation provides in a preferred form of the invention the peptide/fimbrial
35 protein conjugate is subjected to a partial denaturation

prior to admixture with the peptide/fatty acid conjugate.

As will be appreciated by those skilled in the art while any peptide can be used in the vaccine of the first aspect of the present invention the preferred peptides are
5 LHRH derived peptides. Where the peptide is an LHRH peptide, or a derivative, the vaccine may be used for chemical sterilization of animals or for use as an alternative to the present LHRH agonist and antagonist therapy for human sex hormone dependent cancers. Four of
10 the most commonly occurring human sex hormone dependent cancers are prostate, breast, endometrial and ovarian cancer. These conditions may be susceptible to treatment by vaccination with the vaccine of the present invention using the LHRH peptide or derivative. The vaccine of the
15 present invention could also be used as an adjunct to the present LHRH agonist and antagonist therapy for these diseases.

It will be appreciated by those skilled in the art that a number of modifications may be made to the peptides
20 preferably used in the present invention without deleteriously effecting the biological activity of the peptide. This may be achieved by various changes, such as insertions, deletions and substitutions, either conservative or non-conservative in the peptide sequence
25 where such changes do not substantially alter the nature of the immune response raised by the peptide. By conservative substitutions the intended combinations are:-

G,A; V,I,L,M; D,E; N,Q; S,T; K,R,H; and F,Y,W.

The vaccine of the first aspect of the present
30 invention will also have applicability in the treatment of diseases such as AIDS, malaria, influenza, zone pellucida peptide epitopes or hepatitis where peptide epitopes have been identified and a vaccine approach is possible.

The vaccine of the first aspect of the present
35 invention comprising an admixture of peptide/fatty acid.

conjugate with peptide/carrier protein conjugate when used without oil or alum adjuvants induces high level antibody responses in animals. The ability of this vaccine to induce such high level response without requiring oil or alum based adjuvants is of particular benefit. Such findings will allow the use of this vaccine where the use of oil based adjuvants is not permitted and the use of alum based adjuvants is being questioned, e.g., human vaccines or vaccines for companion animals.

10 The preferred form of preparing the peptide/type 4 fimbrial protein conjugate involves using the method disclosed in Australian patent application No 17049/88. By using this method the desired peptide can be expressed in association with Type 4 fimbriae by genetically
15 engineered strains of *Pseudomonas*. This method is, however, to some extent, limited in that the peptide epitope size that can be incorporated into the fimbriae is dependent on the degree of modification which can be made before the fimbriae is no longer properly assembled.
20 Investigations have shown that peptides of up to approximately 20 amino acid residues can be produced in association with the fimbriae in *Pseudomonas aeruginosa*. Accordingly, it is presently preferred that the peptides are able to be expressed in association with fimbriae in
25 *Pseudomonas aeruginosa*. If, however, the peptide epitope is too large or cannot be expressed in this fimbrial system it is still possible to use the vaccine of the present invention by simply chemically conjugating the peptide with the fimbrial protein.

30 The response to the vaccine component of the present invention may also be enhanced by the use of T-cell, H_2 receptor antagonists, e.g., cimetidine or carnosine, or other immunomodulators e.g., cytokines or immunostimulatory peptides which may act as immunomodulators to overcome
35 carrier protein unresponsiveness or to further enhance the

immune system.

The present inventors also believe that the antibody response in an animal to an antigen may be heightened by the use of fimbrial protein of a Type 4 fimbriate bacteria.

5 Accordingly, in a third aspect the present invention consists in a method of enhancing the antibody response to an antigen in an animal, the method comprising to the animal an effective amount of the antigen conjugated to the fimbrial protein of a Type 4 fimbriate bacteria.

10 As is disclosed in application No PCT/AU90/00599 a wide range of fatty acids may be linked to the peptide via a tromethamine or ethanolamine derivative. In addition, 1-3 fatty acids may be linked to the peptide. It should also be noted that the peptide linked to the fatty acids
15 can be of virtually unlimited size.

In order that the nature of the present invention may be more clearly understood a preferred form thereof will now be described with reference to the following examples and figures in which:-

20 Figs. 1-4 show the results of a comparative vaccination trial using ten animals/group, mice were given two vaccinations at four week intervals and data collected four weeks after the second vaccination; (Fig. 1 control, Fig 2 100µg untreated LHRH-fimbriae and Freund's
25 Incomplete Adjuvant (FIA), Fig. 3 Wild Type Fimbriae, Fig. 4 untreated LHRH-Fimbriae admixture with LHRH ala-tris-tripalmitate (ATP3) conjugate;

Fig. 5 shows the antibody response for the animals set out in Figs. 2 and 4, the light hatch represents the
30 serum LHRH antibody levels for group in Fig 2 and the dark hatch group Fig 4. Groups Fig. 1 and Fig. 3 had no LHRH antibodies;

Fig. 6 shows the results of admixture vaccinations using LHRH-acid treated fimbriae or LHRH-ovalbumin
35 conjugate with the ATP3 conjugate. Four groups of ten

mice were given a primary vaccination followed by a secondary vaccination four weeks later.

6(a) mice vaccinated with acid treated LHRH-fimbriae adjuvanted with FIA (ATF-FIA);

5 6(b) mice vaccinated with acid treated LHRH-fimbriae in admixture with ATP3 conjugate (ATF-ATP3);

6(c) mice vaccinated with LHRH-ovalbumin conjugate adjuvanted with FIA (OV-FIA); and

10 6(d) mice vaccinated with LHRH-ovalbumin conjugate in admixture with ATP3 conjugate (OV-ATP3). Testes weights are presented for each individual animal;

Fig. 7 shows the antibody results from the vaccinations set out in Fig. 6 ☐ ATF-FIA; ☒ ATF-ATP3; ☒ OV-FIA; and ☒ OV-ATP3.

15 Fig. 8 shows antibody results obtained in immunocastrated mice after regeneration of testes were revaccinated with untreated LHRH-fimbriae and FIA;

☐ prior to revaccination, ☒ one week after revaccination, ☒ nine weeks after revaccination. The background absorbance level for this data was 0.19 units;

20 Fig. 9 shows the results of mouse vaccination trials using acid treated LHRH-fimbriae adjuvanted with FIA and potentiated with cimetidine (100 μ l of 5mg/ml given every second day for 6 days after vaccination). The mice were vaccinated with 100 μ g of fimbrial protein given

25 subcutaneously and revaccinated four weeks later. The testes weights and LHRH antibody levels for each individual mouse are shown in Figs. 9a and 9b respectively;

Fig. 10 shows the results of mouse vaccination trials

30 using acid treated LHRH-fimbriae, immunopotentiated with carnosine (100 μ l of 5mg/ml given every second day for 6 days after vaccination) and adjuvanted with FIA. The mice were vaccinated with 100 μ g of fimbrial protein given subcutaneously and revaccinated four weeks later. The

35 testes weights and LHRH antibody levels for each

individual mouse are shown in Figs 10a and 10b respectively;

Fig. 11 shows the results of a mouse vaccination trial using acid treated LHRH-fimbriae in a water/glycerol mixture and not adjuvanted with oil or alum. The mice were vaccinated with 150mg of protein subcutaneously and given a booster vaccination four weeks later. The testes weights and LHRH antibody levels for each individual mouse four weeks after the booster vaccination are shown in Figs 11a and 11b respectively; and

Fig. 12 shows the results of mouse vaccination trials using acid treated LHRH-fimbriae in admixture with LHRH-ATP3 and immunopotentiated with either cimetidine or carnosine. The mice were given a primary vaccination with 100mg acid treated LHRH-fimbriae together with 2mg of LHRH-ATP3 and immunopotentiated with three doses of 500mg of either cimetidine or carnosine. A secondary vaccination was given four weeks later, the testes weights and LHRH antibody levels for each individual mouse eight weeks after the primary vaccination is shown in Figs 12a (□ cimetidine, ■ carnosine) and 12b (▨ cimetidine, ▩ carnosine) respectively;

Materials and Methods

Plasmids and bacterial strains

The multifunctional vector pFEM2 was constructed and used for oligonucleotide-directed mutagenesis of the fimbrial subunit gene and for the testing of the fimbrial expression of modified subunits in P.aeruginosa. In brief, the plasmid contains a c1857 gene cartridge obtained from pC121 and a fimbrial subunit expression cartridge from pJSM129. These were recombined to give a vector which can replicate in E.coli and P.aeruginosa, has a M13 intergenic region for the production of single stranded DNA and carries a Dichleobacter nodosus A1 serotype fimbrial subunit expression cartridge. Regulated

expression of the fimbrial subunit is possible because of the plasmid-borne cl857 gene. The P.aeruginosa strain K/2Pfs was used for fimbrial subunit expression.

Competent cells were prepared using the procedure

- 5 described for P.aeruginosa. Transformed cells were propagated at 37°C or at 42°C when fimbrial subunit expression was desired. P.aeruginosa transformants were cultured in Trypticase Soy Broth (TSB, 30 gms/litre) containing 0.15 mg/ml carbenicillin.

10 Expression of Fimbrial Subunit Bearing the LHRH Graft

- Oligoneuclotide directed mutagenesis and hybridisation screening was used to insert the LHRH related peptide into the pFEM/2 plasmid. DNA sequence analysis of a plasmid designated pMF2/1, isolated from one
15 of these clones confirmed that the desired peptide had been grafted to the fimbrial subunit. pMF2/1 was used to transform P.aeruginosa K2 and expression of the modified subunit tested. A protein with a molecular weight greater than that of the unmodified fimbrial subunit was induced
20 and could be detected extracellularly. The extracellular material was immunologically identified as D.nodosus fimbrial subunit and electron microscope examination revealed that the material existed as intact fimbriae. These results showed that fimbrial subunit bearing the
25 LHRH graft was assembled and harvested as morphogenetically correct, extracellular structures.

Induction of Fimbrial Subunit Expression

- For the induction of fimbriae in P.aeruginosa, colonies from 37°C overnight culture plates were mixed
30 into 8ml portions of Trypticase Soy Broth (TSB) and 2ml aliquots spread onto 245mm x 245mm TSB agar plates containing 0.15mg/ml of Carbenicillin. Plates were incubated at 42°C for 18 to 24 hr and cells harvested by scraping into a minimal volume of phosphate-buffered
35 saline. Fimbriae were precipitated from cell-free

- 10 -

supernatants of the harvested cells in the presence of 0.1M magnesium chloride. Whole fimbrial samples were then analyzed for the protein content of samples using a dye-binding procedure (BCA Pierce, USA) and samples taken for the electrophoretic analysis using a urea-SDS-polyacrylamide gel system.

Acid Treatment of PAK2 Fimbriae

Fimbrial preparations after $MgCl_2$ precipitation were washed with distilled water and resuspended at a concentration of 10mg/ml. The pH of the fimbrial solution was adjusted to a pH of 1 using AR grade H_3PO_4 (85% w/v, Ajax Chemicals). The pH adjusted solution was then left undisturbed at $40^\circ C$ for 16 hours. The acid treated fimbriae were then neutralised with 100 μ l of 10 M NaOH and examined using non denaturing PAGE. The protein concentration was determined using a BCA method (Pierce, USA) and then stored at $-20^\circ C$ until required.

Determination of LHRH antibody levels using ELISA.

Samples of blood were taken from mice at death and the sera from each stored at $-20^\circ C$ until the antibody level was determined. A 200 μ l aliquot of gelatin conjugated LHRH at $4^\circ C$. These were then washed twice with phosphate buffered saline containing 0.5ml of Tween 20/litre (PBS-Tween) and blocked for 1.5 hours with 0.5 gms/100 ml gelatin. The plates were washed twice and 200 μ l of 1:2500 mouse sera, from the experimental and control mice, added and left for 1.5 hours. These were again washed and a 1:2000 dilution of (horse radish peroxidase) HRP conjugated rabbit anti-mouse IgG (Dako Corp., Denmark) added to each. These were left for 1.5 hours washed 3 times with PBS-Tween and treated with OPD to determine the response. The absorbances were measured at 492 nm using a plate reader.

Immunization of Mice.

Twenty-five-day-old male Swiss/Balb c mice were

immunized intraperitoneal (I.P.) with 100 μ g to 150 μ g of LHRH engineered PAK2 fimbriae in sterile distilled water at 1:1 (50 μ l : 50 μ l) mixture with Freund's Incomplete Adjuvant (FIA) (Difco, UK) or Titremax (CytRx Corp, USA). A secondary immunisation was given at one month and the animals were sacrificed at two months after the primary immunisation. Blood and testes samples were taken at this point and used for LHRH antibody determination and statistical analysis.

10 Preparation of Ala-T Mono, Di and Tri Palmitate

1 miliequivalent of LHRH peptide was reacted with 3 miliequivalents of O(N-succinimidyl)-N,N,N',N' - tetramethyl uronium tetrafluoro borate (Fluka, 85975) in dimethyl formamide and N-ethyl-di-isopropyl amine was added to the apparent pH of 8.7. Active ester was formed in excess of 75% yield. To this solution 2 miliequivalents of ala-tris(palmitate, dipalmitate or tripalmitate) was added in dichloromethane (2-3ml) and stirred for 1 hour. The apparent pH was readjusted to 8.5.

20 The reaction was monitored by HPLC using mobile phase consisting of

- (i) Buffer A - 0.1% TFA : acetonitrile, 50:50, and
- (ii) Buffer B - acetonitrile: THF, 50:50 using a Novapak C4 column.

25 Purification of the products was by semi-preparative HPLC, on a C4 column and after the removal of solvents lyophilised from t-butyl alcohol.

Results

In order to assess the effectiveness of the LHRH related peptide/fimbrial protein and peptide/fatty acid conjugates mice were immunised with a variety of vaccine combinations. Figures 1-4 shows a comparative vaccination trial using 10 animals per group. Mice were given two vaccinations at a four week interval and data collected four weeks after the second. The control animals (Fig. 1)

- 12 -

and animals which received wild type fimbriae (fimbriae containing no peptide additions) (Fig. 3) showed no reduction in testes weight. (Testes weight of less than 0.25g is considered a positive response) In contrast the
5 animals vaccinated with acid treated LHRH related peptide/fimbrial protein with Freund's Incomplete Adjuvant (FIA) showed a marked reduction in testes weight. Similarly, mice receiving untreated LHRH related
10 peptide/fimbrial protein in admixture with peptide/fatty acid conjugate showed a marked reduction in testes weight. Figure 5 shows the antibody response in these animals.

Figure 6 shows the results of mouse vaccinations with LHRH fimbrial protein conjugate and LHRH-ovalbumin
15 conjugate with ATP3 fatty acid conjugate (ala-tris-tripalmitate). Groups of 10 mice were vaccinated twice at four week intervals with combinations of the ATP3 conjugate and either LHRH conjugated to ovalbumin or LHRH conjugated to acid treated fimbriae.
20 Data was collected four weeks after the second vaccination. The positive Control for the data set was LHRH acid treated fimbriae or LHRH-ovalbumin conjugate with FIA. Both LHRH presenting carrier molecules in combination with the fatty acid peptide conjugate yielded
25 good physical responses, i.e., gonadal atrophy. The antibody responses for these animals are presented in Figure 7.

To exemplify the vaccines reversibility, five mice which had been vaccinated with LHRH-related
30 peptide/fimbrial protein and (FIA) were left to recover after gonadal atrophy had occurred, i.e. 120 days after the primary vaccination. These mice were revaccinated with LHRH-fimbriae and 14 days later they were once again displaying marked gonadal atrophy. This indicates that
35 the castration effect induced by the LHRH/fimbrial antigen

is reversible and can be maintained with subsequent vaccinations. Antibody levels for these mice one week and four weeks after vaccination are presented in Figure 8.

5 Figs. 9 and 10 show that the antibody response and reduction in testes weight can be increased by using the immunopotentiating agents cimetidine and carnosine.

Fig. 11 shows that significant reductions in testes weight and increases in antibody levels can be obtained by administering acid treated LHRH-fimbriae conjugate in the
10 absence of an adjuvanting agent. This ability to enhance antibody response using the Type 4 fimbriae is unexpected. It is believed that this unexpected result opens up the possibility of enhancing antibody responses by using Type 4 fimbriae as a "self-adjuvanting" carrier
15 for peptides conjugated to the fimbriae.

Fig. 12 shows that significant reductions in testes weight and high antibody levels are obtained in animals vaccinated with acid treated LHRH-fimbriae in conjunction with LHRH-ATP3. It is of particular note that these
20 reductions in testes weight and antibody levels were obtained without the use of oil or alum based adjuvants.

This "self-adjuvanting" ability of the vaccine admixture is of particular relevance given the inability to use oil based adjuvants in humans and the questions
25 being raised concerning the use of alum based adjuvants.

As will be seen the present invention provides a vaccine which can be used to raise significant antibody levels against peptides without the use of oil or alum based adjuvants. This discovery is of great utility given
30 the present concern regarding the use of oil and alum based adjuvants. Further, these findings open a new avenue for vaccines for human use.

It will be appreciated by persons skilled in the art that numerous variations and/or modifications may be made
35 to the invention as shown in the specific embodiments

- 14 -

without departing from the spirit or scope of the invention as broadly described. The present embodiments are, therefore, to be considered in all respects as illustrative and not restrictive.

CLAIMS:-

1. A vaccine for use in raising antibodies to a peptide, the vaccine comprising in admixture the peptide conjugated to 1 to 3 fatty acids and the peptide conjugated to a
5 carrier protein.
2. A vaccine as claimed in claim 1 in which the carrier protein is selected from the group consisting of fimbrial subunit protein, ovalbumin, bovine serum albumin, tetanus toxin and keyhole limpet haemocyanin.
- 10 3. A vaccine as claimed in claim 1 or claim 2 in which the carrier protein are fimbrial subunit proteins assembled into mature fimbriae.
4. A vaccine as claimed in claim 3 in which the fimbrial subunit protein is Type 4 fimbriae.
- 15 5. A vaccine as claimed in any one of claims 1 to 4 in which the peptide is conjugated to 1 to 3 fatty acids via a tromethamine derivative or an ethanolamine derivative.
6. A vaccine as claimed in any one of claims 1 to 4 in which the peptide is selected from the group consisting of
20 LHRH peptides, AIDS peptide epitopes, malaria peptide epitopes, influenza peptide epitopes and hepatitis peptide epitopes and zona pellucida peptide epitopes.
7. A vaccine as claimed in claim 6 in which the LHRH peptide is selected from the group consisting of
25 SGGWSYGLRPGG, WSYGLRP, WSYGWLP and WSYGLQP.
8. A vaccine for use in chemical sterilisation, the vaccine comprising a peptide which has or includes an amino acid selected from the group consisting of
30 SGGYSYGLRPGG, WSYGLRP, WSYGWLP and WSYGLQP, the peptide being conjugated either to 1 to 3 fatty acids or a fimbrial subunit protein.
9. A vaccine as claimed in claim 8 in which the fimbrial subunit proteins are assembled into mature fimbriae.
10. A vaccine as claimed in claim 9 in which the fimbrial
35 subunit protein is Type 4 fimbriae.

11. A vaccine as claimed in any one of claims 8 to 10 in which the peptide is conjugated to 1 to 3 fatty acids via a tromethamine derivative or an ethanolamine derivative.
12. A vaccine as claimed in any one of claims 3 to 11 in which the peptide/fimbrial protein conjugate is exposed to an acid treatment at pH less than or equal to 4.
13. A vaccine as claimed in claim 12 in which the peptide/fimbrial protein is exposed to an acid treatment at pH 1.
14. A vaccine as claimed in any one of claims 1 to 13 in which the peptide is linked to 3 fatty acids each of which is the same fatty acid.
15. A vaccine as claimed in any one of claims 1 to 14 in which the fatty acid has a carbon chain of 3 to 18 carbon atoms.
16. A vaccine as claimed in claim 15 in which the fatty acid has a carbon chain of 16 carbon atoms.
17. A vaccine as claimed in any one of claims 1 to 16 in which the vaccine further includes cimetidine, carnosine, cytokines or immunostimulatory peptides.
18. A method of chemically sterilising an animal comprising administering to the animal a vaccine as claimed in any one of claims 3 to 17.
19. A method of treating a subject suffering from prostate cancer, breast cancer, endometrial cancer or ovarian cancer comprising administering to the subject a vaccine as claimed in any one of claims 3 to 17.
20. A method of enhancing the antibody response to an antigen in an animal, the method comprising administering to the animal an effective amount of the antigen conjugated to the fimbrial protein of a Type 4 fimbriate bacteria.
21. A method as claimed in claim 20 in which the antigen is a peptide selected from the group consisting of SGGWSYGLRPGG, WSYGLRP, WSYGWLP and WSYGLQP.

- 17 -

22. A method as claimed in claim 20 or claim 21 in which fimbrial protein is exposed to an acid treatment at pH less than or equal to 4.

23. A method as claimed in claim 22 in which the fimbrial
5 protein is exposed to an acid treatment at pH 1.

1/17

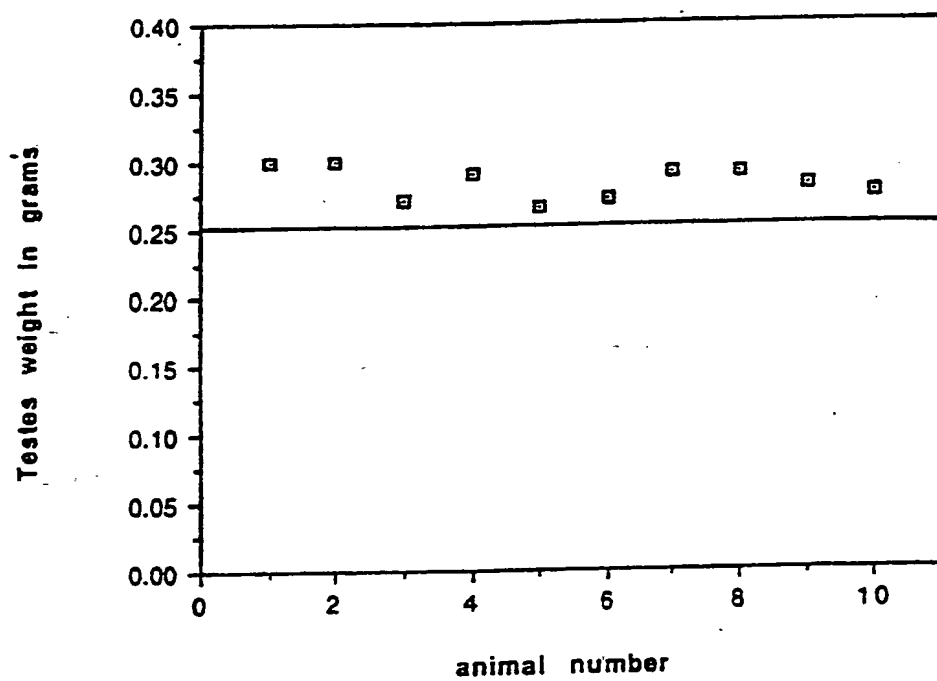


FIG. 1

2/17

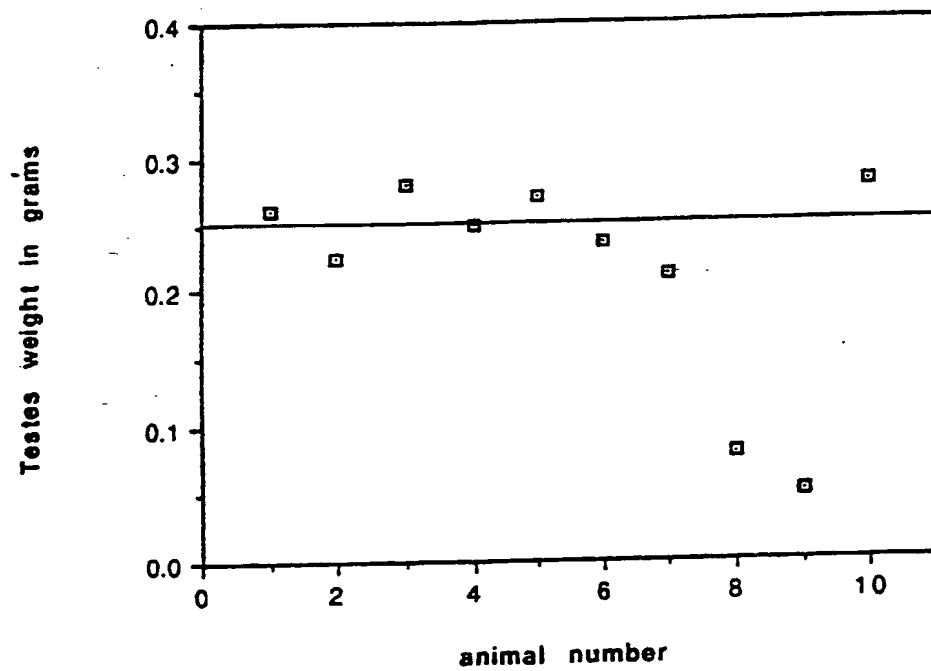


FIG. 2

3/17

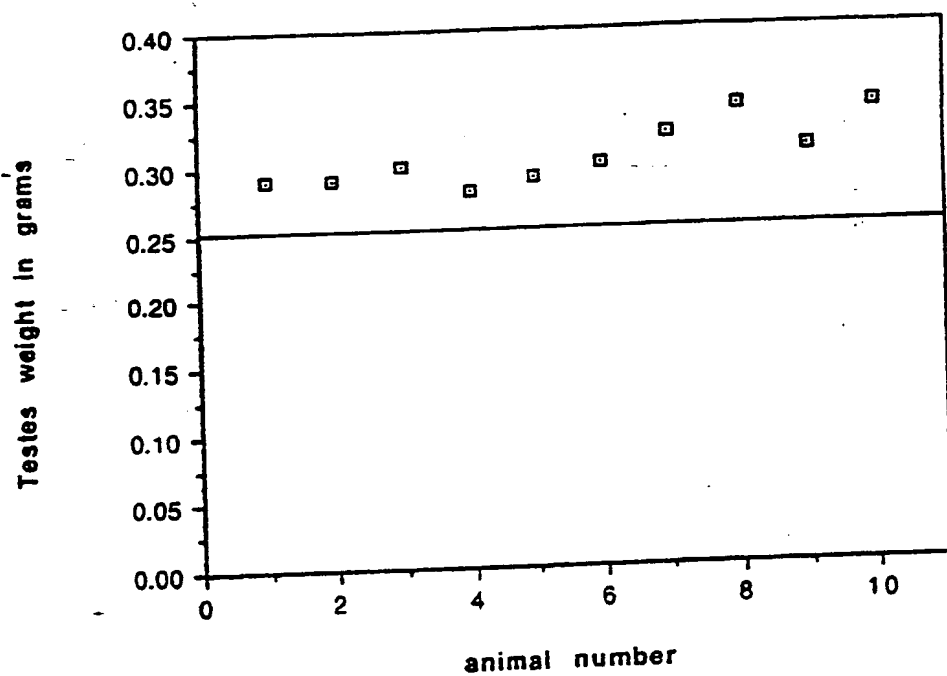


FIG. 3

4/17

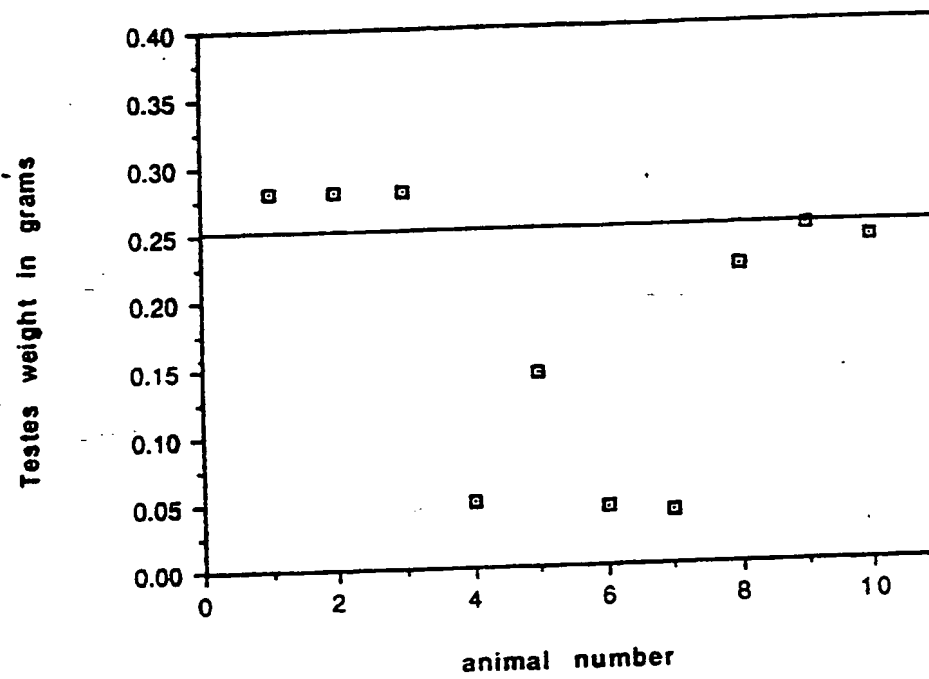


FIG. 4

5/17

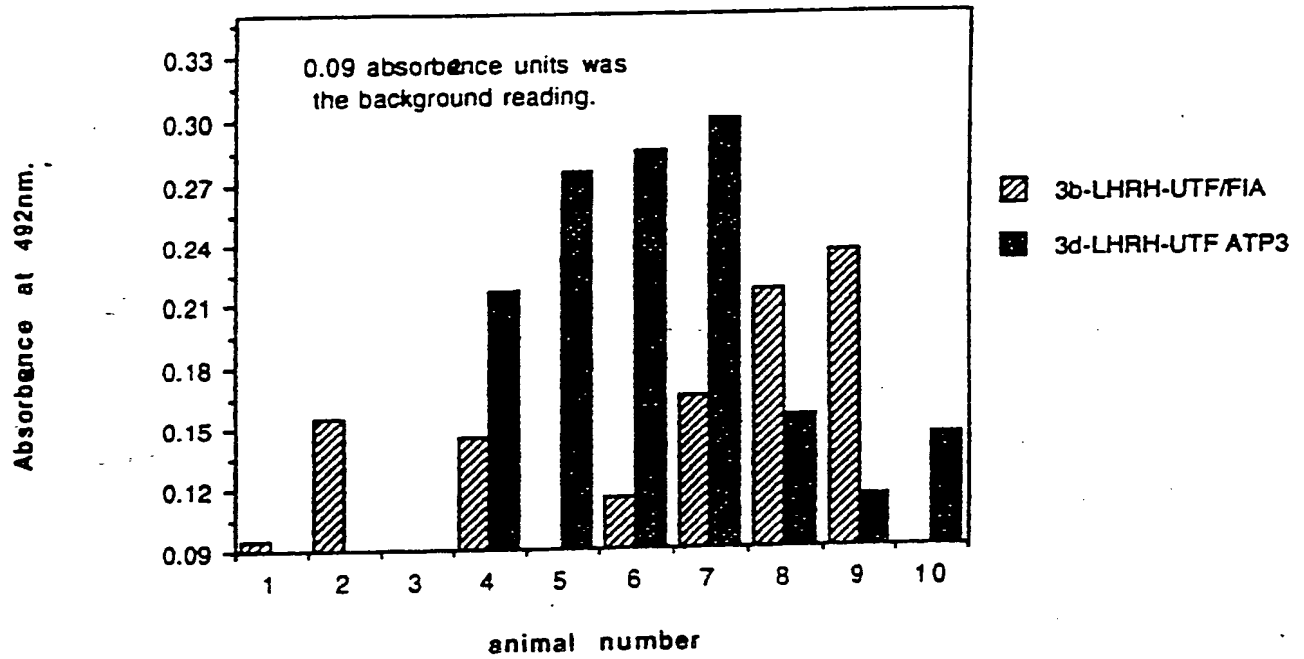


FIG. 5

6/17

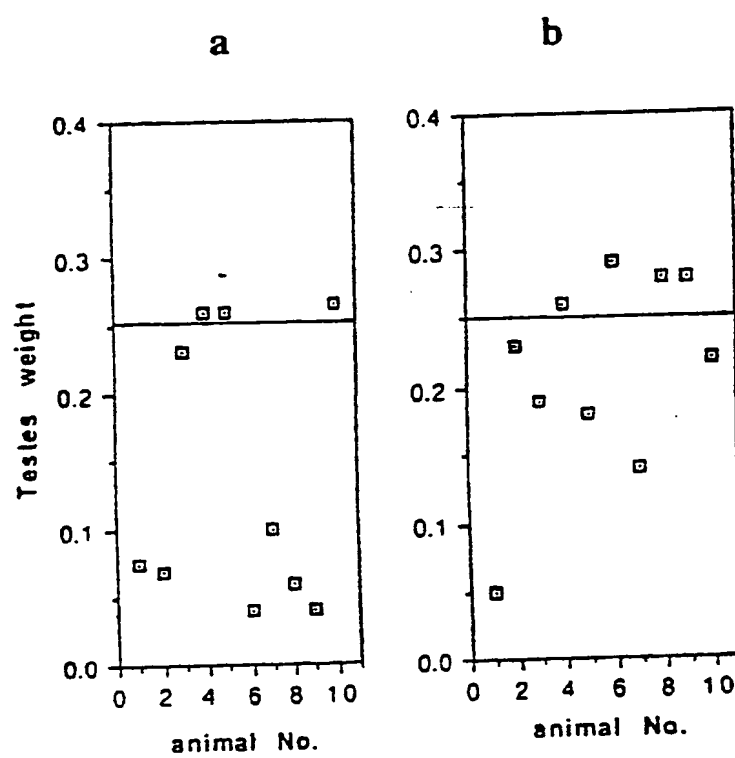


FIG. 6a

FIG. 6b

7/17

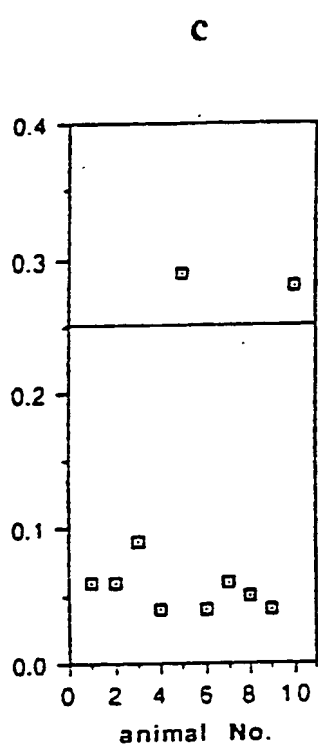


FIG. 6c

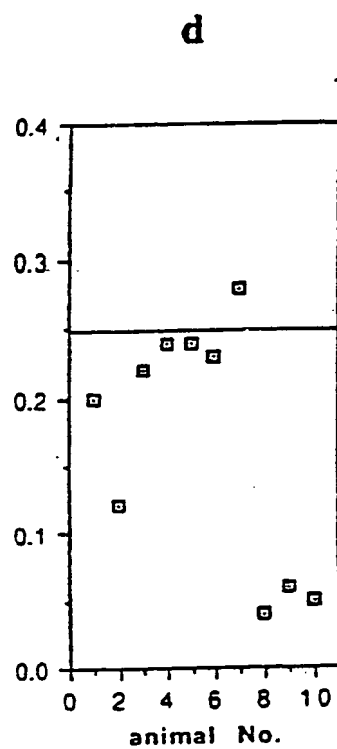


FIG. 6d

8/17

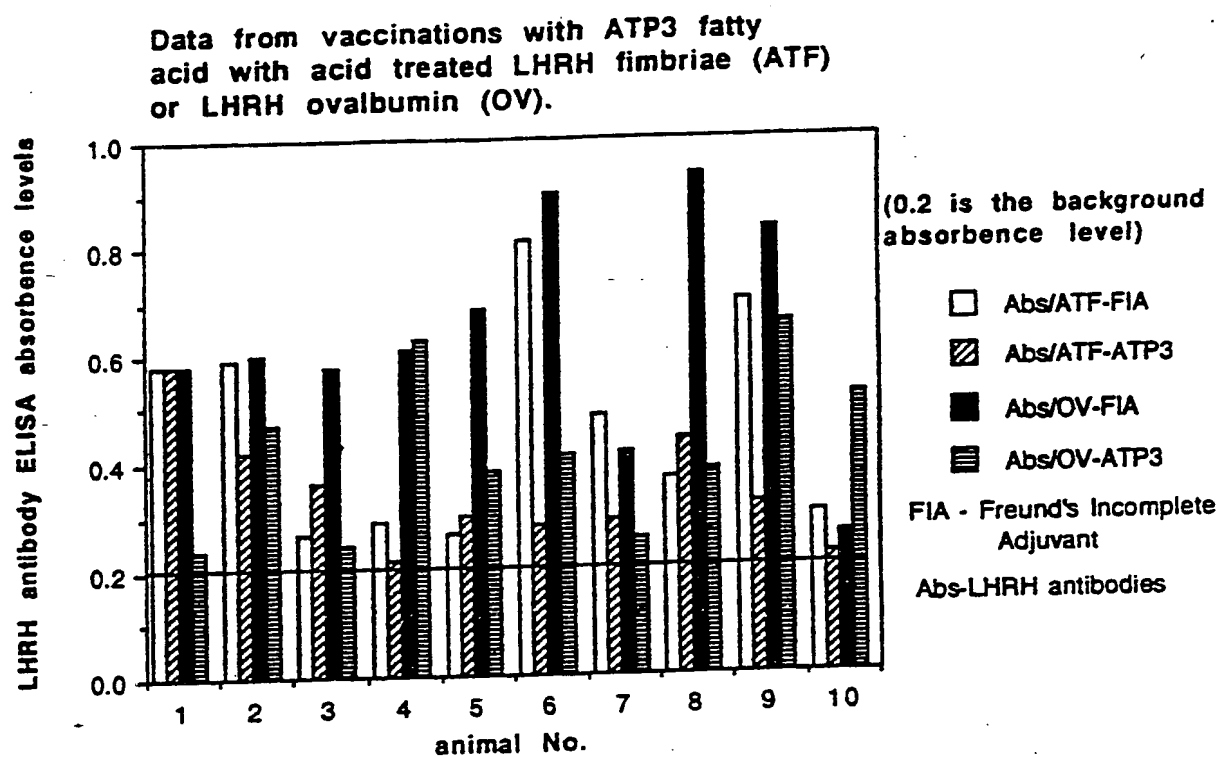


FIG. 7

9/17

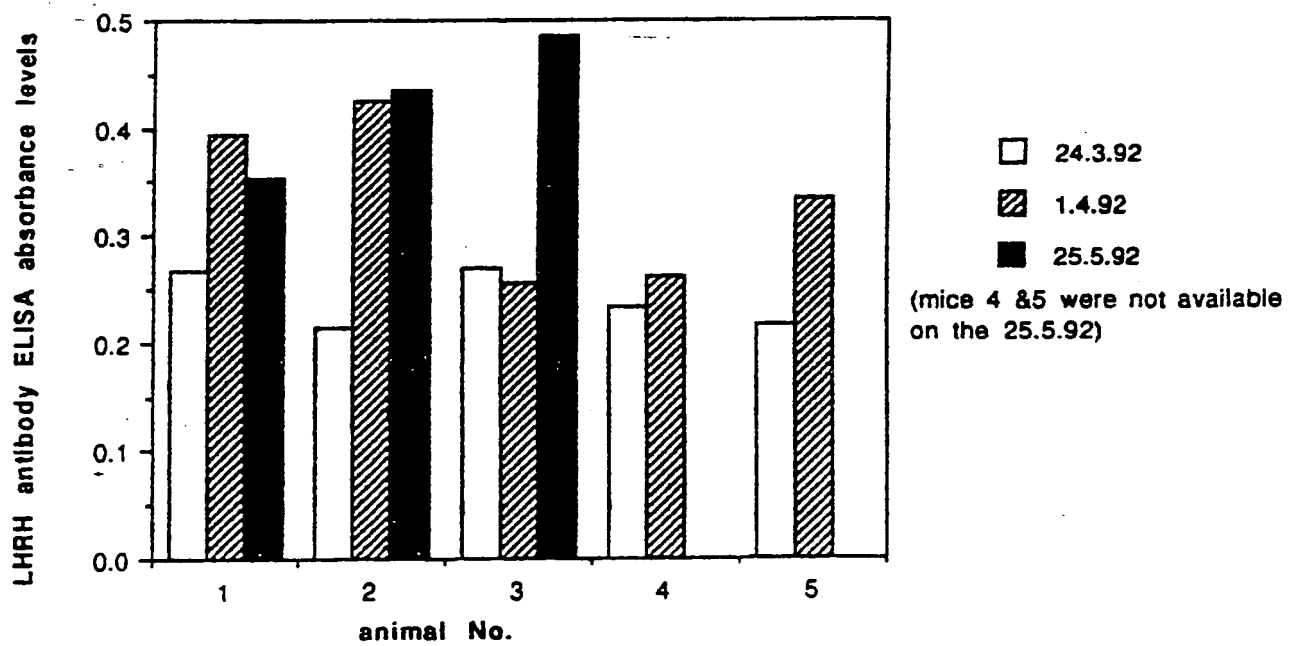


FIG. 8

10/17

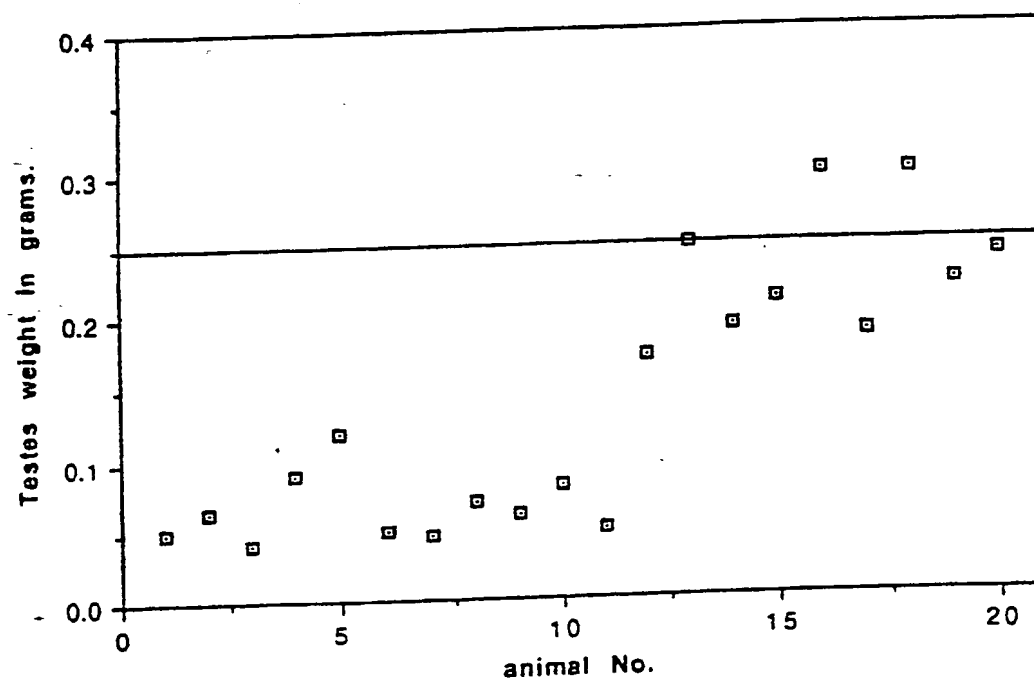


FIG. 9a

11/17

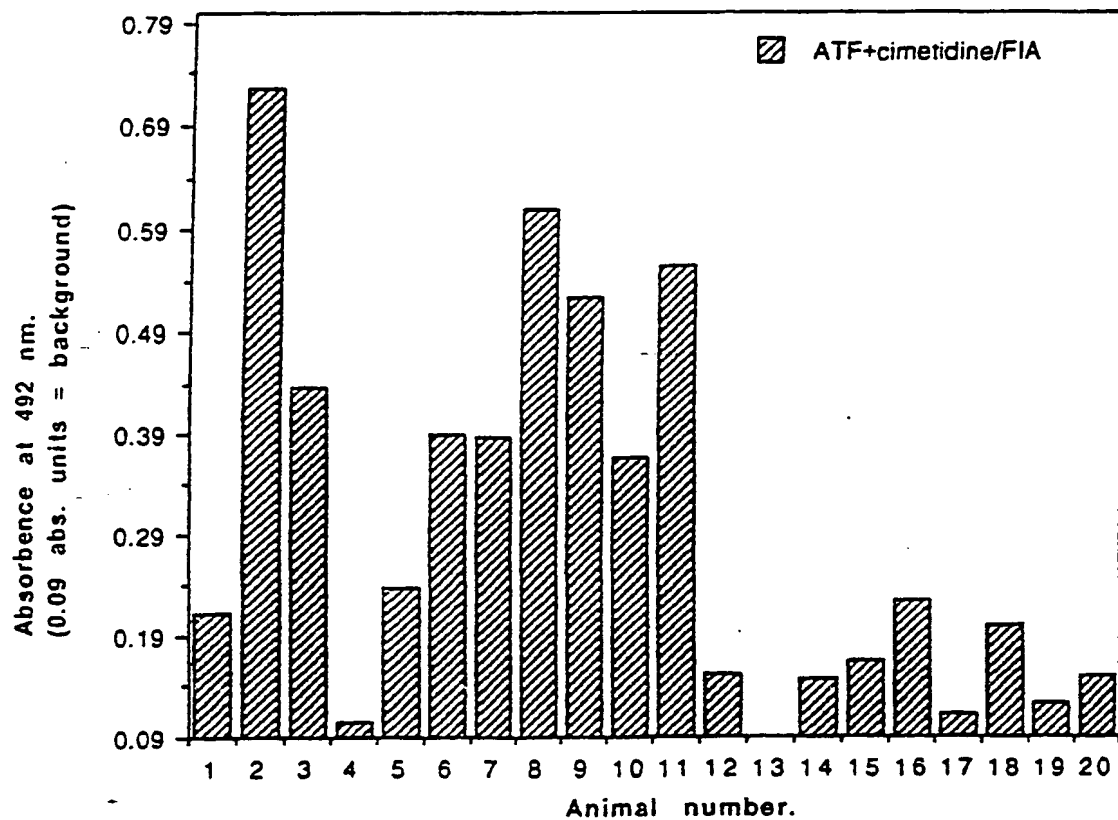


FIG. 9b

12/17

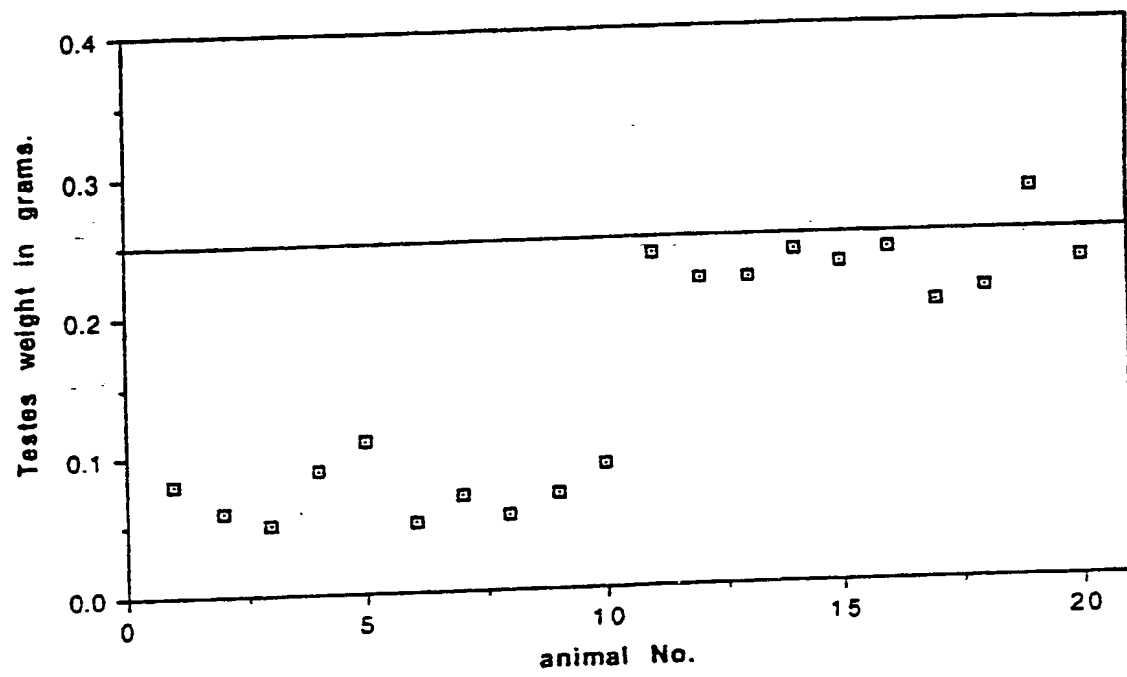


FIG. 10a

13/17

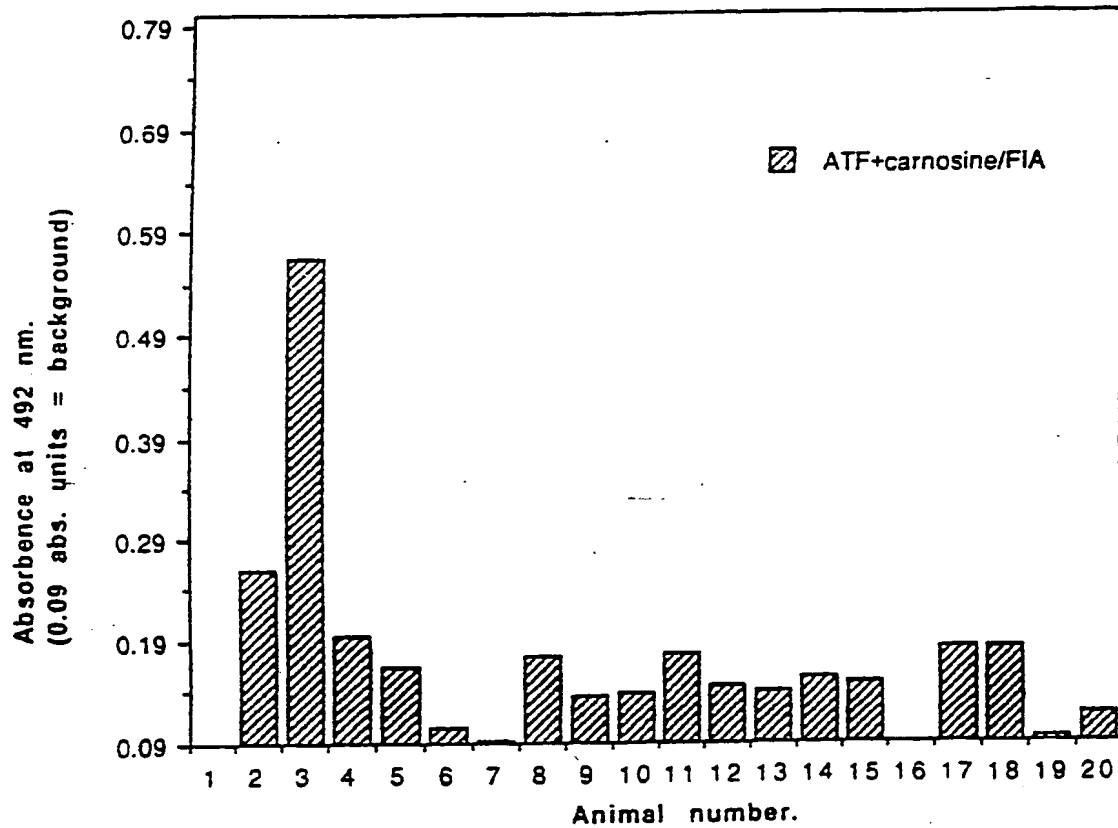


FIG. 10b

14/17

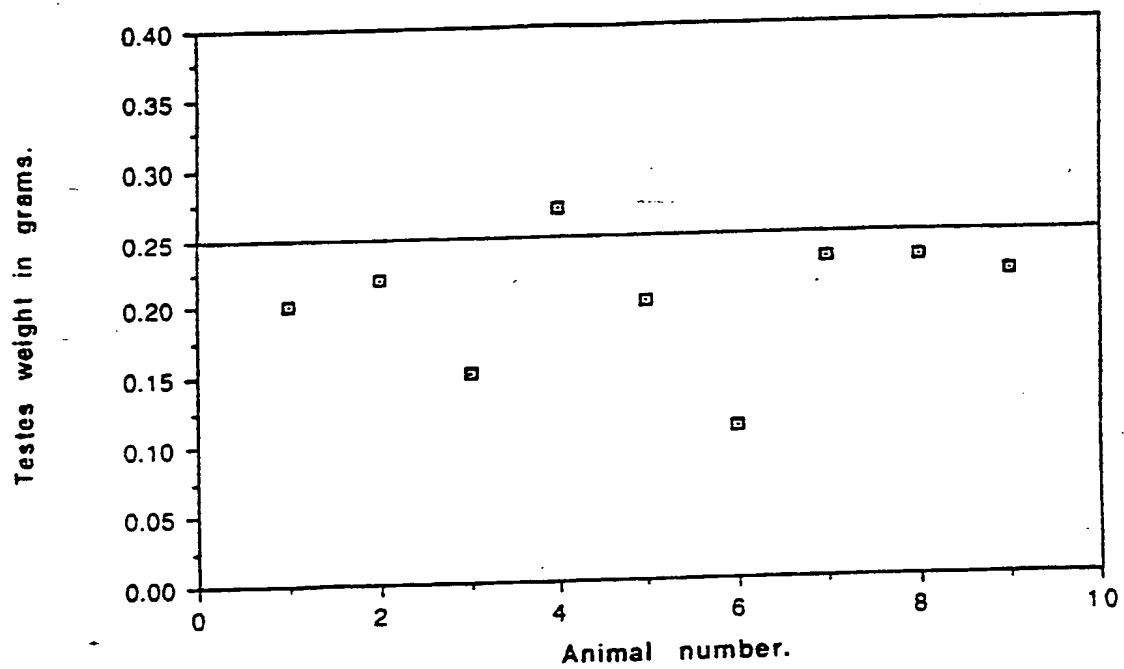


FIG. 11a

15/17

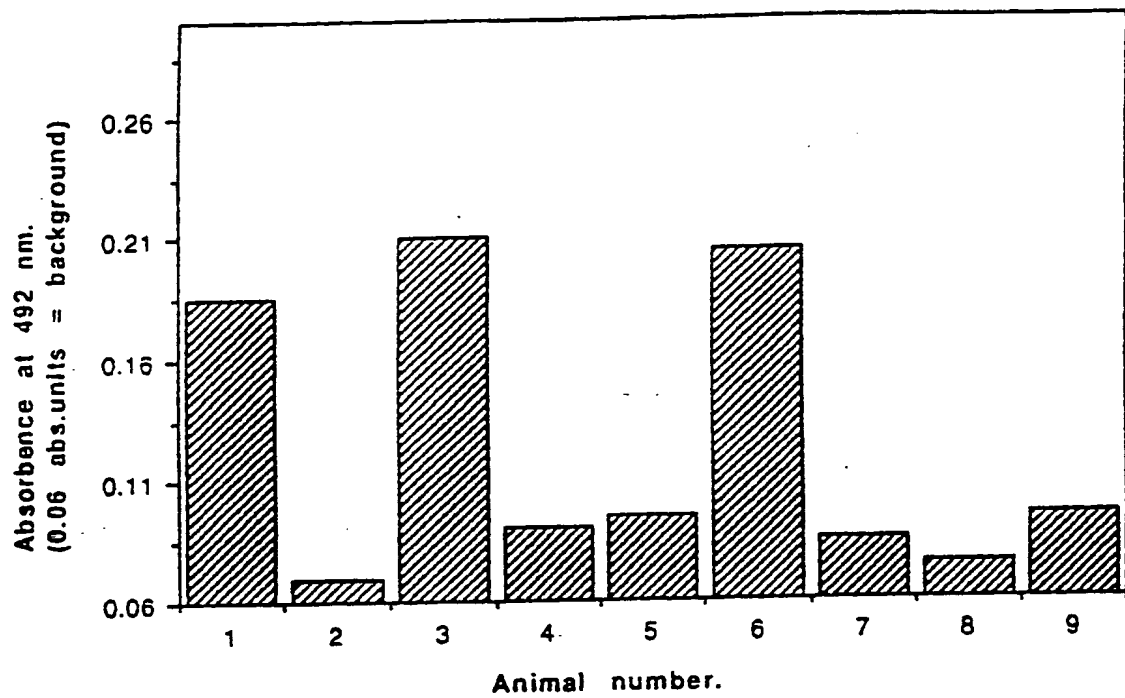


FIG. 11b

16/17

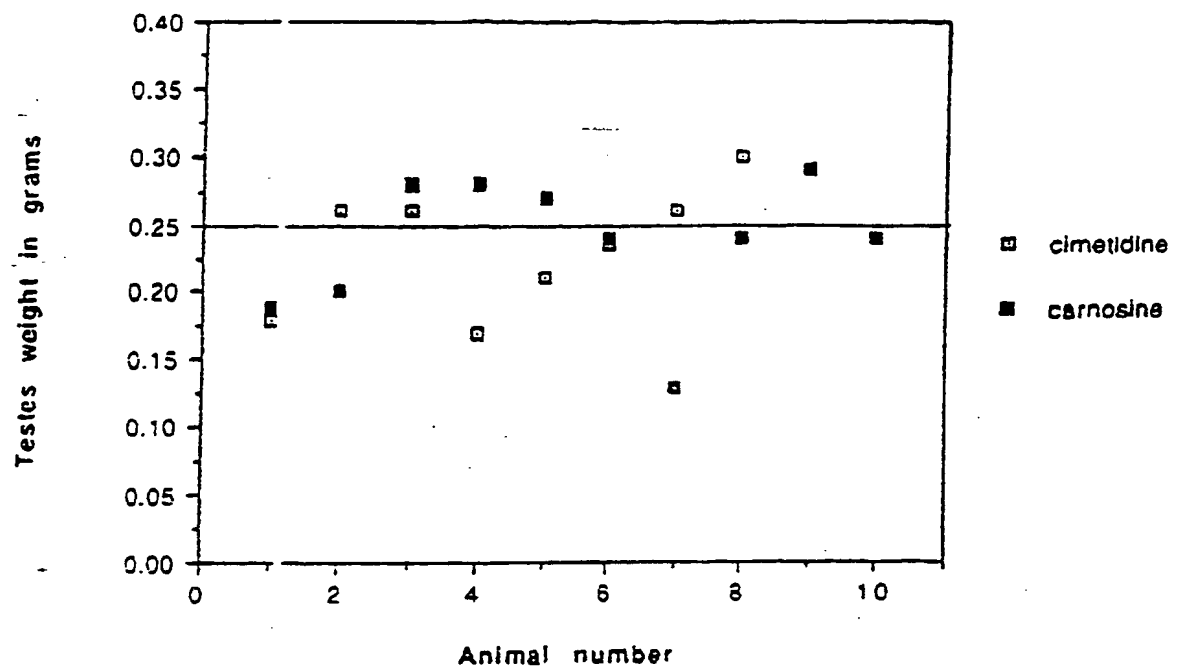


FIG. 12a

17/17

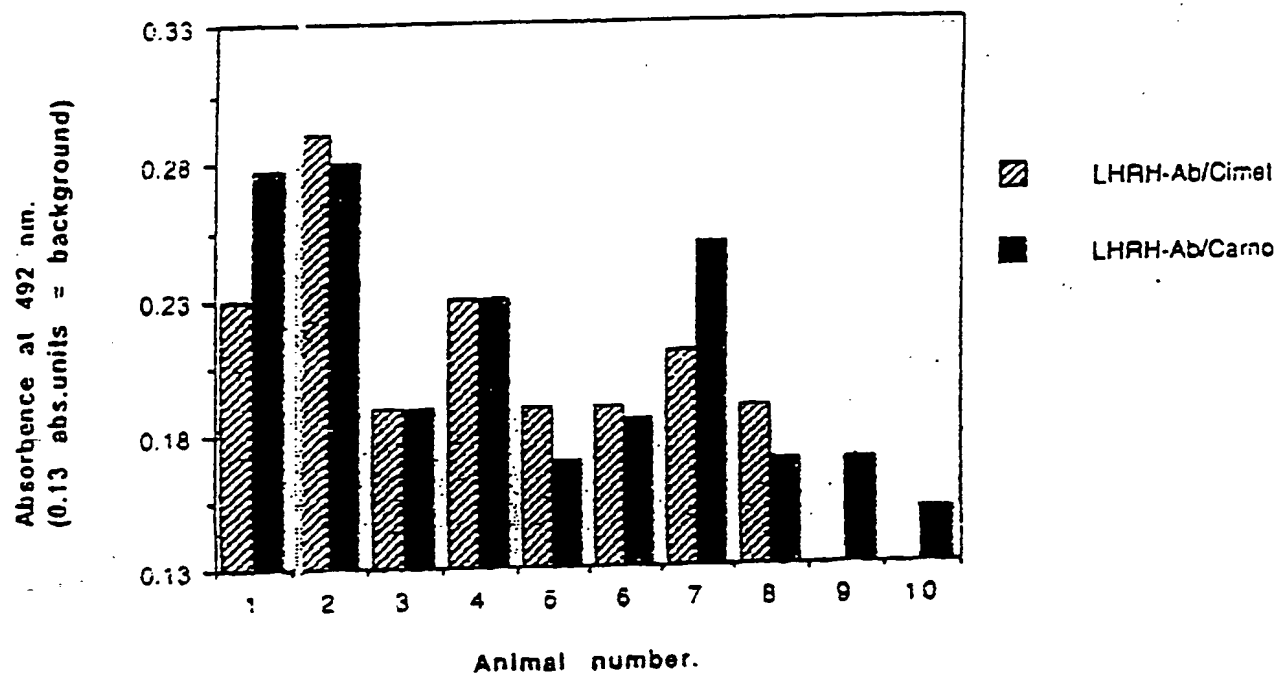



FIG. 12b

A. CLASSIFICATION OF SUBJECT MATTER Int. Cl. ⁵ I.P.C. ⁵ A61K 39/39, A61K 30/00, A61K 39/02 According to International Patent Classification (IPC) or to both national classification and IPC				
B. FIELDS SEARCHED Minimum documentation searched (classification system followed by classification symbols) I.P.C. ⁵ A61K 39/39, A61K 37/43 Documentation searched other than minimum documentation to the extent that such documents are included in the fields searched Electronic data base consulted during the international search (name of data base, and where practicable, search terms used) DERWENT: KEYWORDS: LHRH OR PSEUDOMON OR (TYPEQ4 AND FIMBIA); ADJUVANT CHEM. ABS: KEYWORDS: AS ABOVE				
C. DOCUMENTS CONSIDERED TO BE RELEVANT				
Category *	Citation of document, with indication, where appropriate, of the relevant passages	Relevant to Claim No.		
Y	AU,A,17049/88 (COMMONWEALTH SCIENTIFIC AND INDUSTRIAL RESEARCH ORGANISATION) 3 November 1988 (03.11.88). See pages 16-18.	1-4,6,17,20		
Y	AU,A,11017/88 (COMMONWEALTH SCIENTIFIC AND INDUSTRIAL RESEARCH ORGANISATION) 10 August 1988 (10.08.88). See pages 11-33, claims 5-7,11	1-2,6,18		
Y	AU,A,77584/87 (BUNGE (AUSTRALIA) PTY LTD) 25 February 1988 (25.02.88). See whole document.	1-2,6,18		
<div style="display: flex; justify-content: space-between;"> <div> <input checked="" type="checkbox"/> Further documents are listed in the continuation of Box C. </div> <div> <input checked="" type="checkbox"/> See patent family annex. </div> </div>				
<table style="width: 100%; border: none;"> <tr> <td style="width: 50%; vertical-align: top;"> * Special categories of cited documents : "A" document defining the general state of the art which is not considered to be of particular relevance "E" earlier document but published on or after the international filing date "L" document which may throw doubts on priority claim(s) or which is cited to establish the publication date of another citation or other special reason (as specified) "O" document referring to an oral disclosure, use, exhibition or other means "P" document published prior to the international filing date but later than the priority date claimed </td> <td style="width: 50%; vertical-align: top;"> "T" later document published after the international filing date or priority date and not in conflict with the application but cited to understand the principle or theory underlying the invention "X" document of particular relevance; the claimed invention cannot be considered novel or cannot be considered to involve an inventive step when the document is taken alone "Y" document of particular relevance; the claimed invention cannot be considered to involve an inventive step when the document is combined with one or more other such documents, such combination being obvious to a person skilled in the art "&" document member of the same patent family </td> </tr> </table>			* Special categories of cited documents : "A" document defining the general state of the art which is not considered to be of particular relevance "E" earlier document but published on or after the international filing date "L" document which may throw doubts on priority claim(s) or which is cited to establish the publication date of another citation or other special reason (as specified) "O" document referring to an oral disclosure, use, exhibition or other means "P" document published prior to the international filing date but later than the priority date claimed	"T" later document published after the international filing date or priority date and not in conflict with the application but cited to understand the principle or theory underlying the invention "X" document of particular relevance; the claimed invention cannot be considered novel or cannot be considered to involve an inventive step when the document is taken alone "Y" document of particular relevance; the claimed invention cannot be considered to involve an inventive step when the document is combined with one or more other such documents, such combination being obvious to a person skilled in the art "&" document member of the same patent family
* Special categories of cited documents : "A" document defining the general state of the art which is not considered to be of particular relevance "E" earlier document but published on or after the international filing date "L" document which may throw doubts on priority claim(s) or which is cited to establish the publication date of another citation or other special reason (as specified) "O" document referring to an oral disclosure, use, exhibition or other means "P" document published prior to the international filing date but later than the priority date claimed	"T" later document published after the international filing date or priority date and not in conflict with the application but cited to understand the principle or theory underlying the invention "X" document of particular relevance; the claimed invention cannot be considered novel or cannot be considered to involve an inventive step when the document is taken alone "Y" document of particular relevance; the claimed invention cannot be considered to involve an inventive step when the document is combined with one or more other such documents, such combination being obvious to a person skilled in the art "&" document member of the same patent family			
Date of the actual completion of the international search 30 October 1992 (30.10.92)		Date of mailing of the international search report 5 Nov 1992 (05.11.92)		
Name and mailing address of the ISA/AU AUSTRALIAN PATENT OFFICE PO BOX 200 WODEN ACT 2606 AUSTRALIA Facsimile No. 06 2853929		Authorized officer  John G. Hanson Telephone No. (06) 2832262		

C(Continuation). DOCUMENTS CONSIDERED TO BE RELEVANT		
Category*	Citation of document, with indication, where appropriate of the relevant passages	Relevant to Claim No.
E, Y	DE, A1, 4003944 (PRINZHAUS, GERHARD) EARLIEST PRIORITY DATE 14 FEBRUARY 1990 (14.02.90) See abstract	1-2, 6, 19
Y	AU, A, 42186/89 (PITMAN-MOORE, INC) 5 April 1990 (05.4.90) See pages 12-14	1-2, 6, 19
Y	US 5017558 (VYAS) 21 May 1991 (21.06.91). See column 6, lines 49-58, column 6, lines 68 to column 7, line 5	1-2, 6, 17
Y	AU, A, 3446/89 (SYNPHARM LTD AND SANTER, VIVIEN, BEDFORD) 2 November 1989 (02.11.89) See column 5, page 6, claim 4.	1-2, 6, 17-20
Y	AU, A, 22755/88 (UNIVERSITY OF SASKATCHEWAN) 20 April 1989 (20.04.89). See examples.	1-2, 6, 18

[illegible]

